1 and 2 is striking. An understanding would involve at a minimum some knowledge of metal-ligand bond strengths in complexes 2, 3, 6, and 7.

Most studies of C-H activation have focussed on aromatic and aliphatic hydrocarbons. In the equally important activation of olefins, the relative stability of  $L_nM(H)(CH=CH_2)$  and  $L_nM(\eta^2-C_2H_4)$  is a central question.

Acknowledgment. We gratefully acknowledge the support of this work by the Natural Sciences and Engineering Research Council and thank Professor Helmut Werner for helpful discussions.

## Protonated Nitric Acid. Experimental Evidence for the Existence of Two Isomers

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Relatively little is known about the positive ion chemistry of  $HNO_3$ , e.g., its proton affinity (PA) is reported to be intermediate between those of  $H_2O$  and  $NH_3$ , which amounts to an uncertainty of almost 40 kcal mol<sup>-1</sup>.<sup>1</sup> We have investigated by Fourier-transform (FT-ICR) and chemical ionization (Cl) mass spectrometry, using  $H_3^+$ ,  $CH_5^+$ , and  $H_3O^+$  as the ionic reactants, the process

$$HNO_3 + AH^+ \rightarrow H_2NO_3^+ + A \tag{1}$$

previously studied in a flowing afterglow.<sup>2,3</sup> Occurrence of reaction 1 has unequivocally been established by triple-resonance ICR experiments, demonstrating, in addition, slow decomposition of 1 into NO<sub>2</sub><sup>+</sup> and NO<sup>+</sup>.<sup>4</sup> The CH<sub>4</sub>/CI spectrum of aqueous HNO<sub>3</sub> displays 1 as the predominant peak, together with its hydrate, NO<sub>2</sub><sup>+</sup>, and NO<sup>+</sup> (Figure 1).<sup>5</sup> Since the basicity of HNO<sub>3</sub> is hardly accessible to equilibrium measurements owing to the decomposition of 1 under ICR conditions, we resorted to the less reliable "bracketing" technique,<sup>1</sup> in experiments carried out either on isolated ions by FT-ICR spectrometry or by ClMS. The basicity of HNO<sub>3</sub> falls between those of H<sub>2</sub>O and of CF<sub>3</sub>COOH (or CF<sub>3</sub>CH<sub>2</sub>OH), leading to an estimated PA(HNO<sub>3</sub>) = 168 ± 2 kcal mol<sup>-1</sup>, not inconsistent with recent theoretical results.<sup>6</sup> From the PA value, one can derive  $\Delta H_f^{\circ}$  (H<sub>2</sub>NO<sub>3</sub><sup>+</sup>)  $\approx$  166 kcal



Figure 1. Typical CH<sub>4</sub>/Cl spectrum of 68% aqueous HNO<sub>3</sub>, recorded at ca. 1 Torr, source temperature 40 °C, by using a 5982A Hewlett-Packard quadrupole spectrometer.



Figure 2. (A) Metastable peak due to the loss of water from ions of type II prepared according to process 1,  $A = CH_4$ , in methane at ca. 0.1 Torr. (B) Same peak from ions of type I, obtained by process 2,  $A = CH_4$ , in methane at ca. 1 Torr.

mol<sup>-1</sup>, hence  $D(NO_2^+-H_2O) \approx 10$  kcal mol<sup>-1</sup>, and  $-\Delta H_{(1)} \approx 67$ (A = H<sub>2</sub>), 36 (A = CH<sub>4</sub>), and  $\approx 2$  (A = H<sub>2</sub>O) kcal mol<sup>-1</sup>.<sup>1</sup> We have exploited as well another long-known<sup>7</sup> route to 1:

$$C_2H_5ONO_2 + AH^+ \rightarrow H_2NO_3^+ + C_2H_4 + A$$
 (2)

a process exothermic by ca. 18 (A = CH<sub>4</sub>) and 50 (A = H<sub>2</sub>) kcal mol<sup>-1</sup>.

Structurally diagnostic techniques provide strong and mutually supporting evidence for the existence of two isomers of 1, isomer 1 being detectable in the ionic populations from reactions of low exothermicity, i.e., from (1),  $A = H_2O$ , and (2),  $A = CH_4$ , while isomer 11 is detectable only as a product from highly exothermic processes, i.e., from (1),  $A = H_2 \text{ or } CH_4$ , and (2),  $A = H_2$ , under conditions of inefficient collisional deactivation.<sup>8</sup> Structural discrimination between 1 and 11 is based on the following evidence.

<sup>(1)</sup> All thermochemical data are taken from the compilation of Lias et al. (Lias, S. G.; Liebman, J. F.; Levin, R. D. J. Phys. Chem. Ref. Data 1984, 13, 695).

<sup>(2)</sup> Fehsenfeld, F. C.; Ferguson, E. E. J. Chem. Phys. 1973, 59, 6272.
(3) Fehsenfeld, F. C.; Howard, C. J.; Schmeltekopf, A. L. J. Chem. Phys. 1975, 63, 2835.

<sup>(4)</sup> The ICR spectra were recorded with a Nicolet FTMS 1000 instrument, measuring the sample pressure with a Granville-Phillips 280 Bayard-Alpert ion gauge, at a typical resolution of 10<sup>3</sup> fwhh at mass 100, with a trapping voltage of 1.0 V, electron-beam energy 15 eV, under a total pressure of ca.  $4 \times 10^{-7}$  Torr.

<sup>(5)</sup> H<sup>+</sup>(HNO<sub>3</sub>)<sub>x</sub>(H<sub>2</sub>O)<sub>y</sub> clusters have been reported, see: Kay, B. D.; Hermann, V.; Castleman, A. W., Jr. *Chem. Phys. Lett.* **1981**, 80, 469 and references therein.

<sup>(6)</sup> According to  $6-31G^{**}//44-31G$  SCF calculations by Nguyen et al. (Nguyen, M. T.; Hegarty, A. F. J. Chem. Soc., Perkin Trans. 2 1984, 2043), the PA of HNO<sub>3</sub> exceeds that of H<sub>2</sub>O by ca. 1 kcal mol<sup>-1</sup>. Such results, while internally consistent, overestimate absolute basicities, e.g., the calculated PA(H<sub>2</sub>O) exceeds the experimental value by some 13 kcal mol<sup>-1</sup>, which could affect the calculated  $D(NO_2^*-H_2O)$  energy.

<sup>(7)</sup> Nixon, W. B.; Bursey, M. M. Tetrahedron 1970, 50, 4389.

<sup>(8)</sup> These observations do not imply that II is the only or the most abundant isomer formed whenever the protonation process is highly exothermic but simply that only under the specified set of conditions it becomes detectable by MIKE and CID spectrometry. Such structurally diagnostic techniques detect the fraction of the ions that undergo unimolecular or collisionally induced decomposition in the appropriate regions of the spectrometer, rather than sampling the relative ionic abundances in the ion source.

(i) Irrespective of their source, metastable decomposition of ions 11 into  $NO_2^+$  and  $H_2O$  occurs with a large release of kinetic energy (747  $\pm$  10 meV), giving a typical, dish-topped peak (Figure 2A), while ions I decompose giving an non-Gaussian peak with a much lower kinetic energy release (Figure 2B). Such a difference is particularly significant, in that kinetic energy release is primarily dependent on the reverse activation energy not on the internal energy of the decomposing ions.<sup>9</sup> Large releases are typical of rearrangements, while simple bond cleavages have little or no reverse activation energies.<sup>10</sup> It follows that II must undergo rearrangement prior to metastable H<sub>2</sub>O loss, which is not the case of 1.

(ii) The collisional-induced dissociation (CID) spectra, depurated from the unimolecular contribution, are also largely different, the C1D spectrum of 1 displaying three fragments, i.e.,  $H_2O^+$ ,  $NO^+$ , and  $NO_2^+$ , in the approximate 0.7:1:9 ratio, while that of 11 displays only NO<sup>+</sup> and NO<sub>2</sub><sup>+</sup> in the approximate 1:2 ratio.

(iii) Further structural discrimination is provided by the different metastable decomposition of the ions from the reaction

$$H_2NO_3^+ + H_2^{18}O \rightarrow H_2NO_2^{18}O^+ + H_2O$$
 (3)

unequivocally established by FT-ICR spectrometry and occurring as well under Cl conditions.<sup>11</sup> lons l from (3) undergo metastable decomposition into unlabeled  $NO_2^+$ , losing exclusively  $H_2^{18}O_1$ , while labeled ions 11 undergo nearly statistical (typically 2.5:1) metastable loss of  $H_2O$  and of  $H_2^{18}O.^{12}$ 

Overall, the above results suggest that 1 is the more stable isomer, characterized by a hydrated nitronium ion structure, whose metastable loss of water involves a simple bond cleavage, without a large release of kinetic energy. The presence of a discrete  $H_2O$ 

$$H_{2}O \xrightarrow{+}{N} \stackrel{+}{<} O \xrightarrow{HO} \xrightarrow{+}{N} \stackrel{+}{\rightarrow} O$$

moiety accords well with the selective loss of  $H_2^{18}O$  in the metastable decomposition of ions 1 from process (3), that amounts, in this case, to a simple ligand exchange. The features of ions 11 are consistent instead with a structure containing two OH groups, whose metastable decomposition into  $NO_2^+$  and  $H_2O$ presupposes molecular reorganization, which justifies the large release of kinetic energy. The mixed isotopic composition of water from the metastable decomposition of <sup>18</sup>O-labeled ions 11 accords well with a structure containing no O atom in a preexistent H-O-H group, whose loss can occur preferentially, as from 1, via a simple bond cleavage, requiring no preliminary rearrangement.

Our conclusions qualitatively agree with the results of SCF calculations, which identify a  $H_2O-NO_2^+$  structure as the most stable among the isomers investigated, showing that the relative energies of ions akin to 11 are higher by at least 8 kcal mol<sup>-1,6</sup>

It should lastly be noted that the kinetic energy release from 11 sets a lower limit of the order of 20 kcal mol<sup>-1</sup> to the activation energy for the hydration of  $NO_2^+$  yielding 11 and that there are reasons to believe that the free energy of activation for the  $II \rightarrow$ l isomerization is correspondingly large. Remarkably, the process does not appear to be catalyzed by interaction of 11 with a water molecule.

Acknowledgment. This work has financially been supported by Ministero della Pubblica Istruzione. We are grateful to P. Giacomello for active interest and useful suggestions and to the FT-ICR service of Area della Ricerca di Roma, CNR, Italy.

## Measurement of <sup>1</sup>H-<sup>1</sup>H Coupling Constants in Oligonucleotides by 2D NMR: Application of $\omega_1$ -Decoupling

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Three-bond <sup>1</sup>H–<sup>1</sup>H coupling constants constitute the basic input for the determination of conformation of d-ribose rings in oligonucleotides by NMR spectroscopy.<sup>1</sup> Different techniques, such as  $\omega_1$ -scaled COSY,<sup>2,3</sup> P.E.COSY,<sup>4</sup> and DISCO,<sup>5</sup> to name a few, have been used to derive this structural information.<sup>6-9</sup> While all these have proved extremely useful, any procedure that enhances resolution in the spectra is always a welcome addition. In this context  $\omega_1$ -decoupling in correlated spectroscopy<sup>10,11</sup> is an attractive proposition since this allows complete elimination of multiplicity along the  $\omega_1$ -axis of the two-dimensional spectrum. In this communication we use the following experimental scheme.<sup>11</sup>

$$20 - (\Delta + t_1)/2 - 180 - (\Delta - t_1)/2 - 90 - acquire(t_2)$$

 $t_1$  and  $t_2$  are the usual evolution and detection periods, and  $\Delta$  is a constant time period. In this scheme the heteronuclear H-P coupling will however be retained along the  $\omega_1$ -axis of the twodimensional spectrum. The cross peaks can be phased to pure absorption along both axes, while diagonal peaks will have disperse character along  $\omega_2$  and absorptive character along  $\omega_1$ .

Figure 1 shows the (H1')-(H2', H2") cross peak region of the  $\omega_1$ -decoupled COSY spectrum of the oligonucleotide hairpin d(C-G-C-G-A-G-T-T-G-T-C-G-C-G). It is seen that all the expected cross peaks are present. Both positive (+) and negative (-) signals have been plotted, and the multiplet pattern along  $\omega_2$ is + - + - for H1'-H2" cross peaks and + + - for H1'-H2" cross peaks. In the case of nucleotide units G2, G4, G6, G9, and G12, strong coupling artefacts are seen between the H1'-H2' and H1'-H2" cross peaks. In Figure 2 a particular horizontal slice through the spectrum in Figure 1 is shown in order to bring out the characteristics of the spectrum. The central + and - signals have lower intensities compared to the two outer signals; the intensity difference being more pronounced in G6. This is a consequence of (i) partial cancellation of the + - intensite is in the center and (ii) strong coupling effects in G6 because of which the central two lines have lower intensities inherently. All the cross peak multiplets have been simulated (illustrated on the side in Figure 1) to obtain true peak positions which directly yielded the H1'-H2' and H1'-H2" coupling constants in all units except where strong coupling effects were observed. In the strongly coupled cases, the simulated peak positions do not yield directly the coupling constant values.

- Govil, G.; Hosur, R. V. Conformation of Biological Molecules: New Results from NMR; Springer-Verlag: New York, 1982.
   Hosur, R. V.; Chary, K. V. R.; Ravikumar, M. Chem. Phys. Lett. 1985,
- 116, 105.
- (3) Hosur, R. V.; Chary, K. V. R.; Sheth, A.; Govil, G.; Miles, H. T. J. Biosci. 1988, 13, 71.
  - (4) Muller, L. J. Magn. Reson. 1987, 72, 191.

- (7) Chazin, W. T.; Wuthrich, K.; Hybert, S.; Rance, M.; William, A. D.;
   Leupin, M. J. Mol. Biol. 1986, 190, 439.
   (8) Zhou, N.; Manogaran, S.; Zon, G.; James, T. L. Biochemistry 1988,
- 27, 6013.
- (9) Bax, A.; Lerner, L. J. Magn. Reson. 1988, 79, 429.
  (10) Bax, A.; Freeman, R. J. Magn. Reson. 1981, 44, 542.
  (11) Rance, M.; Wagner, G.; Sorenson, O. W.; Wuthrich, K.; Ernst, R. R. J. Magn. Reson. 1984, 59, 250.

<sup>(9)</sup> Holmes, J. L.; Terlouw, J. K. Org. Mass Spectrom. **1980**, 15, 393. (10) Cooks. R. G.; Beynon, J. H.; Caprioli, R. M.; Lester, G. R. Meta-stable lons; Elsevier: Amsterdam, 1973. (11) Even under Cl conditions,  $H_2NO_2^{18}O^+$  ions arise exclusively from exchange 3, since no appreciable hydration of  $NO_2^+$  has been detected in specific control oversimption.

specific control experiments

<sup>(12)</sup> The MIKE and C1D spectra were recorded by using a ZAB-2F instrument (Micromass, Ltd.), operating the CI source at 160 °C. The spectra represent an average of 100 scans, with an energy resolution of 1.2 eV main-beam width.

<sup>(5)</sup> Oschkinat, H.; Freeman, R. J. Magn. Reson. 1984, 60, 164.
(6) Chary, K. V. R.; Hosur, R. V.; Govil, G.; Chen, C.-q.; Miles, H. T.